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Fish & Wildlife Division

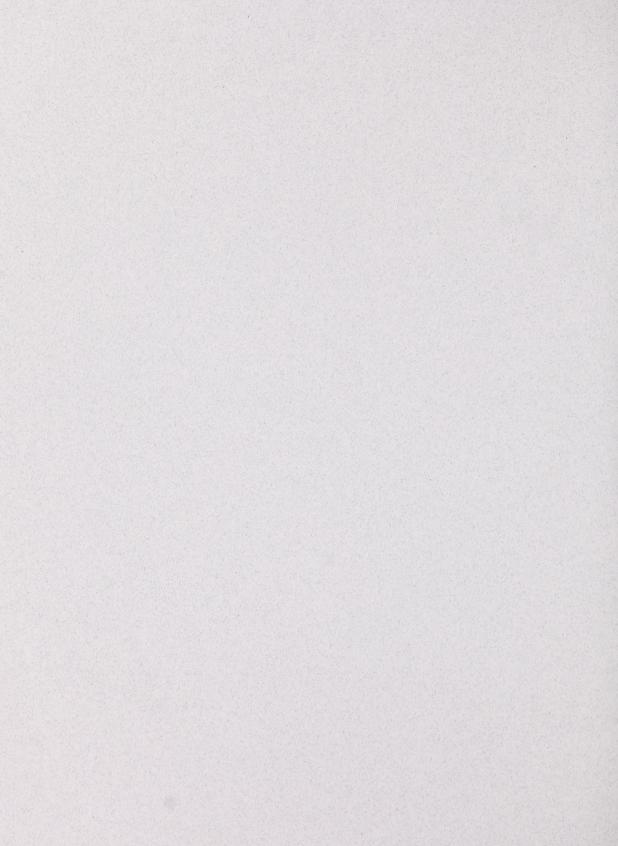
RESOURCE DATA AND SPECIES AT RISK SECTION

RESEARCHING AMPHIBIAN NUMBERS IN ALBERTA (RANA): 2003 PROVINCIAL REPORT



Alberta Species at Risk Report No. 83





Researching Amphibian Numbers in Alberta (RANA)

2003 Provincial Report

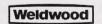
Lisa Wilkinson and Kent Kempin

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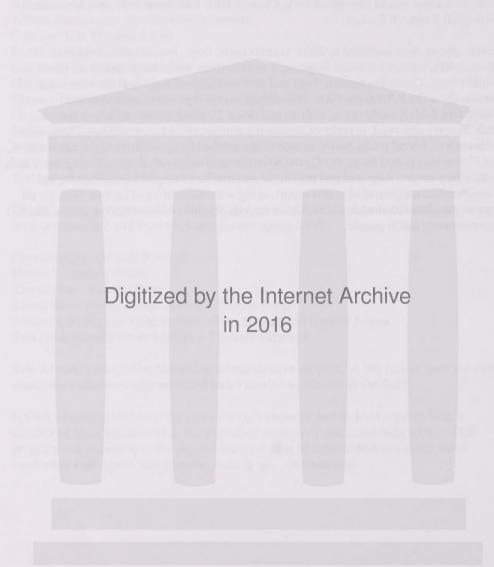
Kris Kendell provided technical and administrative support. A big thanks goes out to the numerous volunteers who provided much needed assistance in the field.

RANA has succeeded over the years through financial and in-kind support from a number of agencies, as well as the continued support of dedicated individuals. This program is a testament to the possibilities and kind of success that can occur when supportive individuals and agencies believe in a common goal.

EXECUTIVE SUMMARY

Six RANA (Researching Amphibian Numbers in Alberta) sites were operated in 2003: Kananaskis (Bow Valley), Hinton (Athabasca Valley), Lesser Slave Lake Provincial Park, Saskatoon Island Provincial Park, Cypress Hills Provincial Park, and Meanook Biological Research Station.

Seven species were captured in 2003: boreal chorus frogs, boreal toads, long-toed salamanders, northern leopard frogs, tiger salamanders, wood frogs, and Columbia spotted frogs. Columbia spotted frogs and northern leopard frogs had not been captured previously in the RANA program. Shoreline pond surveys were conducted at 214 ponds within five RANA study areas, with an additional 29 pond surveys in Jasper National Park. Species observed, in order of decreasing abundance, were: wood frogs, long-toed salamanders, boreal toads, boreal chorus frogs, northern leopard frogs, tiger salamanders, and Columbia spotted frogs. No Canadian toads were observed, despite having study sites within their range. Regional and provincial summaries of amphibian morphological characteristics are provided in this report, along with summaries of capture rates for all species encountered at each of the RANA trapping ponds. Educational programs reached approximately 6,000 people.



1.0 INTRODUCTION

Field data collection for the Researching Amphibian Numbers in Alberta (RANA) program was initiated in 1997 in response to the global decline of amphibians. RANA has two primary objectives: 1) collect long-term data on amphibian species populations in Alberta, and 2) provide public education on the importance of amphibians and wetland conservation. Two sites were operated in 1997, and since that time, an additional five monitoring sites were established, although not all sites can be operated every year.

There were six active sites in 2003: Lesser Slave Lake Provincial Park (est. 1997), Saskatoon Island Provincial Park (est. 1999), Meanook Research Station (est. 1997), Cypress Hills Provincial Park (est. 1999), Athabasca Valley (Hinton; est. 2000), and Bow Valley (Kananaskis; est. 1998). Beaverhill Lake is no longer operated as a RANA site due to logistical constraints.

The following species have been captured or observed in the RANA program: wood frogs (*Rana sylvatica*), boreal chorus frogs (*Pseudacris maculata*), Columbia spotted frogs (*Rana luteiventris*), northern leopard frogs (*Rana pipiens*), boreal toads (*Bufo boreas*), Canadian toads (*Bufo hemiophrys*), long-toed salamanders (*Ambystoma macrodactylum*), and tiger salamanders (*Ambystoma tigrinum*). The only two amphibian species that have not been observed during the RANA program are the great plains toad (*Bufo cognatus*) and the plains spadefoot toad (*Spea bombifrons*), both of which are found in the grassland natural region (Alberta Environmental Protection 1994), which is not currently represented. Only one Canadian toad has been captured since the establishment of the RANA program, in 1998 at Lesser Slave Lake.

Few of Alberta's amphibians are considered 'secure'. Of the species encountered in the RANA program, the northern leopard frog is 'threatened', the Canadian toad is 'data deficient' ('may be at risk'), and the long-toed salamander and Columbia spotted frog are 'sensitive' (Alberta Sustainable Resource Development 2000). Long-toed salamanders are the focal species in Hinton and Kananaskis due to the current provincial objective to monitor their population distribution and trends for at least five years. Additional details of the salamander program are in Wilkinson and Hanus (2003).

Public education has always been a fundamental component of the RANA program. Education takes the form of presentations, public event displays, guided hikes, school talks and other activities. Technical presentations are given to land-use managers (industry and government) and at biological conferences. In conjunction with RANA educational programs, the Alberta Amphibian Monitoring Program (AAMP) is promoted. This program encourages members of the public to record and submit amphibian observations throughout the province. An instruction manual, including identification photos and a tape of frog and toad calls, is also provided to interested individuals.

This document is a compilation of 2003 field results from each RANA site summarizing key findings. Individual field summary reports for 2003 have been developed for Hinton (Hanus 2004), Kananaskis (Rose 2004), Lesser Slave Lake Provincial Park (Kempin 2004), Saskatoon Island Provincial Park (Augustyn 2004), Cypress Hills Provincial Park (Rempel 2004), and Meanook Biological Research Station (Kempin 2004).

2.0 STUDY AREA

The RANA program has sites distributed across Alberta (Figure 1) to represent a variety of natural regions (Alberta Environmental Protection 1994). Cypress Hills is located in a unique montane ecosystem within the grassland natural region, Kananaskis (Bow Valley) is located in the rocky mountain and foothills natural regions, the Hinton (Athabasca Valley) site is located in the foothills natural region, and the Meanook Biological Research Station, Lesser Slave Lake and Saskatoon Island Provincial Park sites are located in different parts of the boreal forest natural region. In addition, pond surveys were conducted in and around Jasper National Park, as part of the long-toed salamander monitoring program.

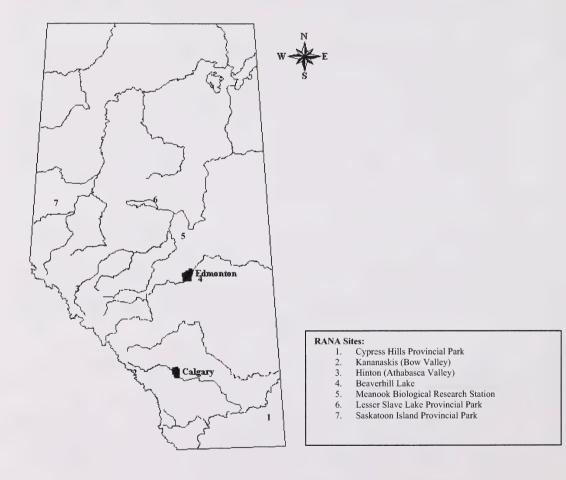


Figure 1. Alberta distribution of RANA sites.

3.0 METHODS

Methods for surveying and capturing long-toed salamanders and other amphibians followed Pretzlaw *et al.* (2002), and are summarized below. Any variations to the methods are included in the individual site reports.

3.1 Pitfall Trapping

In each RANA survey area, there is one pond set up for pitfall trapping. The pond is completely or partially encircled with drift net fencing (Figure 2A). Pitfall traps are placed at 10 m intervals on either side of the fence. Theoretically, amphibians travelling to or from the pond to breed will be intercepted by the fence and travel along the barrier until they fall into a trap. Traps consist of two 6-inch plastic flower pots taped together, creating a 12-inch deep trap (Figure 2B). Traps are covered by a square of coroplast, a sturdy and water resistant plastic, held above the traps by 6-inch nails. Trap covers ensure that amphibians are protected from excessive exposure to sun, which can lead to desiccation, and prevent flooding from rainfall. The following items are placed in each trap: a sponge and/or moss, which is refreshed regularly to retain moisture; a rock that serves as a perch or hiding place; and a stick protruding past the top of the trap to allow small mammals to escape.

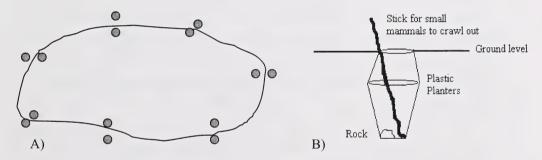


Figure 2. Layout of fencing and pitfall traps (A), and close-up of pitfall trap design (B).

Trapping schedules varied slightly between sites, but traps were generally set during the spring breeding season (May to early June), and late summer (August to early September) when young-of-the-year disperse from the ponds. The Meanook site was operated from July to mid-August for logistic reasons. Throughout the trapping sessions, traps were checked daily or every other day. Traps were generally closed during mid-summer when amphibian movement to and from ponds declined. Captured animals were identified to species, measured, weighed, and sex and age determinations were made when possible. Animals were released on the opposite side of the fence from which they were captured. Other morphological characteristics, such as presence of dorsal stripes and malformations were also recorded. Small individuals captured at the beginning of the season were presumed to be juveniles (young-of-the-year from the previous season). Age and sex differentiation was not possible for all species. Researchers used latex gloves when handling amphibians to avoid contaminating amphibians with oils or hand lotion. Animals being released away from the pond were moistened and placed in a sheltered

location; extra care was taken on hot days, releasing amphibians early or late in the day to prevent desiccation. Environmental data were also collected, including air and water temperature, pond pH, water depth, and other ambient conditions. At the Hinton RANA site, long-toed salamanders were marked with elastomer (a non-toxic latex) as part of an ongoing mark-recapture study (refer to Wilkinson and Hanus 2003 for details).

At the end of each trapping session all traps were closed by either pushing the coroplast cover flush to the ground and covering it with dirt and a rock, or by inverting traps, to ensure animals could not enter. Also, sections of fence were opened at several locations around the pond to allow animals to pass through.

3.2 Shoreline Pond Surveys

Pond surveys were conducted to identify presence of breeding amphibians through observations of eggs, larvae/tadpoles, young-of-the-year, or adults, as well as to hear vocalizations in the spring. Survey efforts were most intensive during spring egg-laying, although some ponds were visited later in the summer to look for presence of larvae and tadpoles.

Surveys were conducted by walking along the perimeter of a pond, or using transects for large and/or partially inaccessible water bodies, and looking for signs of amphibian presence. Surveys could not be conducted under excessively windy or rainy conditions because the surface of the water was disturbed. Similarly, care was taken not to disturb the surface of the water around the shoreline. Data recorded included number of salamander eggs, number of frog egg masses, and number of toad egg strings. For observations of larvae and tadpoles a specific count was made if possible, otherwise qualitative observations were made using estimates of few, moderate, or abundant (a similar system applies to vocalizations). Observations of adults were also recorded. Environmental data were collected as per pitfall trapping ponds, and GPS locations were recorded for all ponds surveyed.

At some sites, the length of survey distance was recorded, allowing for future calculations of number of eggs/meter, enabling relative abundance comparisons over time.

3.3 Reports and Data Storage

In addition to this summary report, a report was produced for each RANA site and distributed to local government offices, stakeholders, funding agencies, participants and volunteers, and other interested parties. All data have been electronically recorded and incorporated into the provincial government Biodiversity/Species Observation Database (BSOD).

4.0 RESULTS

4.1 Pitfall Trapping

Results from spring and late summer pitfall trapping sessions are provided in Table 1 and 2, respectively. In addition, amphibian capture data from each RANA site since their establishment is provided in Appendix 1.

4.1.1 Spring Pitfall Trapping Session

Five of the six active RANA sites were open during the spring trapping session in 2003, and a total of five amphibian species were captured (Table 1). Wood frogs were the most numerous amphibian species captured during the spring trap session (287), with nearly 80% of those captures at Lesser Slave Lake Provincial Park. Long-toed salamanders were the second-most captured species in the spring (218) and were found only at the Hinton and Kananaskis sites, followed by boreal chorus frogs (49), boreal toads (36), and tiger salamanders (6). No one species was present at all trapping sites, but wood frogs were caught at every site except Cypress Hills Provincial Park.

4.1.2 Late Summer Pitfall Trapping Session

Five of the six active RANA sites were open during the late summer trapping session, with Meanook being operational for the early part of this period (July 4 – Aug. 12). A total of seven amphibian species were captured (Table 2). Of the species observed, wood frogs were the most numerous, primarily due to the large number of young-of-the-year captured at the Meanook site (1989). Other amphibian species captured, in decreasing order of abundance, were long-toed salamanders (173), boreal toads (91), northern leopard frogs (71), boreal chorus frogs (32), tiger salamanders (4), and Columbia spotted frogs (1;).

4.2 Shoreline Pond Surveys

Pond surveys were conducted at a total of 216 ponds in Cypress Hills, Hinton, Kananaskis, Lesser Slave Lake, and Saskatoon Island (Table 3), with an additional 29 ponds in Jasper National Park. Wood frogs were the most ubiquitous species, observed at 49% of ponds surveyed. The highest amphibian species diversity, with four species, was shared between the Hinton, Kananaskis, and Saskatoon Island Provincial Park sites. In Cypress Hills, northern leopard frogs were present at 47% of ponds surveyed.

4.3 Education

Education took a variety of forms, including presentations, public displays, guided talks, children's activities, and newspaper/newsletter articles. Site specific amphibian presentations were developed for each RANA site, and posters of local amphibians were also created for some sites. An estimated total of 5969 people were informed about amphibians, RANA, the AAMP, and wetland conservation throughout the province (Table 4). Kananaskis had an active outreach program, using public events to reach large numbers of people, while Cypress Hills used a variety of media to reach people within and around the park. Saskatoon Island also reached a larger number of people through interpretative talks, and school talks were the focus of Hinton and Lesser Slave Lake educational programs. Details about presentations and activities are included in each RANA site report.

Table 1. Spring amphibian captures and morphological characteristics from Alberta RANA sites in 2003.

* Age** Total# Trap Sca Adult 1 0.002 0 1 Adult 24 0.052 10 14 Juvenile 2 0.004 0 0 Overall 26 0.005 10 14 Overall 26 0.006 0 0 Overall 126 0.016 49 76 Overall 27 0.048 4 0 Overall 28 0.068 0 1 Juvenile 25 0.045 0 0 Overall 24 0.05 0 0 Overall 38 0.068 0 1 Overall 24 0.05 0 0 Overall 38 0.068 0 1 Overall 27 0.048 4 0 Overall 39 0.05 0 0 Overall 30 0.00 0 Overall 30 0.00 0 Overall 36 0.01 5 8 Adult 218 0.067 87 129 Overall 6 0.002 0 0 Overall 6 0.002 0 0 Overall 6 0.002 0 0 Overall 21 0.05 3 1 Adult 218 0.067 87 129 Overall 6 0.002 0 0 Overall 27 0.008 19 17					# Per	Cox			Snout-to-Vent	3-Vent	Mass (e	
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BOTO					Night	M	F	Unk.	Mean	Range	Mean	Range
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WOFR Adult 24 0.052 10 14 WOFR Juvenile 2 0.004 0 0 WOFR Overall 26 0.052 10 14 BCFR Overall 2 0.005 0 0 ITSA Overall 6 0.006 0 0 NLFR Overall 126 0.16 49 76 WOFR Overall 156 0.02 6 2 WOFR Juvenile 25 0.045 0 0 WOFR Juvenile 25 0.045 0 0 WOFR Juvenile 25 0.045 0 0 WOFR Overall 21 0.05 3 1 BCFR Overall 36 0.015 6 0 WOFR Overall 36 0.015 6 0 BCFR Overall 36 0.015 6 <th< th=""><th>Hinton</th><th>LTSA</th><th>Adult</th><th>92</th><th>0.19</th><th>38</th><th>53</th><th>1</th><th>6.4</th><th>5.0-7.5</th><th>4.8</th><th>3.0-7.7</th></th<>	Hinton	LTSA	Adult	92	0.19	38	53	1	6.4	5.0-7.5	4.8	3.0-7.7
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NLFR Overall 0 0 0 0	May 16 - June 30	TISA	Overall	9	900.0	0	0	9	9.4	8.5-9.9	28.4	21.6-32.1
BOTO	(1000 trap nights)	NLFR	Overall	0	0	0	0	0	0	0	0	0
LTSA Overall 126 0.16 49 76 WOFR Overall 16 0.02 6 2	Kananaskis	BOTO	Adult	8	0.01	1	7	0	8.9	8.7-0.9	31.7	20.4-47.3
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WOFR Overall 224 0.4 0 0 BCFR Overall 9 0.02 0 0 WOFR Overall 21 0.05 3 1 BCFR Overall 49 0.015 0 1 BOTO Overall 36 0.01 5 8 LTSA Adult 218 0.067 87 129 NLFR Overall 6 0 0 0 TISA Overall 6 0.002 0 0 WOFR Adult 260 0.08 19 17 WOFR Juvenile 27 0.008 0 0	(560 trap nights)	WOFR	Juvenile	25	0.045	0	0	25	2.4	2.0-2.7	1.2	0.5-1.5
BCFR Overall 9 0.02 0 0 WOFR Overall 21 0.05 3 1 BCFR Overall 49 0.015 0 1 BOTO Overall 36 0.01 5 8 LTSA Adult 218 0.067 87 129 NLFR Overall 6 0 0 0 WOFR Adult 260 0.008 19 17 WOFR Juvenile 27 0.008 0 0 WOFR Juvenile 27 0.008 0 0		WOFR	Overall	224	0.4	0	0	224	3.6	2.0-6.4	6.7	0.5-27.0
WOFR Overall 21 0.05 3 1 BCFR Overall 49 0.015 0 1 BOTO Overall 36 0.01 5 8 LTSA Adult 218 0.067 87 129 NLFR Overall 0 0 0 0 TISA Overall 6 0.002 0 0 WOFR Adult 260 0.08 19 17 WOFR Juvenile 27 0.008 0 0	Saskatoon Island	BCFR	Overall	6	0.02	0	0	6	2.5	2.0-2.8	3	2.2-4.7
BCFR Overall 49 0.015 0 1	Apr. 29 - May 19	WOFR	Overall	21	0.05	3	-	17	4.7	3.8-5.7	9.2	5.0-19.0
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BOTO Overall 36 0.01 5 8	Totals	BCFR	Overall	49	0.015	0	1	48	2.3	1.5-3.2	1.5	0.5-4.7
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NLFR Overall 0 0 0 0 0 0 0 Overall 0 0 0 0 0 Overall 0 0 0 0 0 0 0 Overall 0 0 0 0 0 0 0 0 Overall 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0		LTSA	Adult	218	0.067	87	129	2	6.5	4.2-8.1	5.3	3.0-8.5
Overall 6 0.002 0 0 R Adult 260 0.08 19 17 R Juvenile 27 0.008 0 0	(3232 trap nights)	NLFR	Overall	0	0	0	0	0	0	0	0	0
Adult 260 0.08 19 17 Juvenile 27 0.008 0 0		TISA	Overall	9	0.002	0	0	9	9.4	8.5-9.9	28.4	21.6-32.1
Juvenile 27 0.008 0 0		WOFR	Adult	260	80.0	19	17	224	4.4	2.0-6.4	6	5.0-27.0
10 000 10		WOFR	Juvenile	27	0.008	0	0	27	2	1.2-2.7	1.4	0.5-2.1
Overall 28/ 0.09 19 1/		WOFR	Overall	287	0.09	19	17	251	4.3	1.2-6.4	8.7	0.5-27.0

^{*}BCFR (Boreal Chorus Frog), BOTO (Boreal Toad), LTSA (Long-Toed Salamander), WOFR (Wood Frog), NLFR (Northern Leopard Frog), TISA (Tiger Salamander).

^{**}Adult LTSA and WOFR were 2.0g or more, whereas Juvenile LTSA and WOFR were <2.0g.

Table 2. Late summer amphibian captures and morphological characteristics from RANA sites in 2003.

DAMA CITE	Crossos	********	Total#	# Per		Sex		Snout	Snout-to-Vent	Σ	Mass (g)
NAMA SHIE	Species	280	LOTAIN	Night	Σ	ī	Ink	Mean	Range	Mean	Range
	ROTO	Overall	0	0.00	C		7	19	4 0-8 2	96	0 59-5 2
	LTSA	Adult	9	0.01	4	2	0	6.4	5.6-7.5	5.6	3.8-8.4
Hinton	LTSA	YOY	10	0.02	0	0	10	4.1	3.7-4.5	1.7	1.2-2.0
Aug. 11 - Sept 13	LTSA	Overall	16	0.03	4	2	10	5.1	3.7-7.5	3.5	1.2-8.4
(450 trap nights)	WOFR	Adult	21	0.047	7	9	8	4.7	4.2-5.5	13	1.7-18.2
	WOFR	YOY	7	0.015	0	0	7	2.3	1.5-2.6	1.7	1.4-2.0
	WOFR	Overall	28	90.0	7	9	15	4.1	1.5-5.5	10.2	1.4-18.2
Kananaskis	BOTO	Overall	19	0.06	0	0	19	5.7	2.0-11.4	18	0.9-38.3
Aug 17 - Sept 13	CSFR	Overall	1	0.003	0	0	_	8.1	8.1	69.4	69.4
(304 trap nights)	LTSA	Overall	157	0.052	0	0	157	3	1.8-6.9	0.98	0.3-7.0
	WOFR	Overall	202	99.0	0	0	202	1.8	1.4-4.9	92.0	0.3-14.1
	BCFR	Overall	14	0.02	0	0	14	2	1.0-3.6	8.0	0.2-2.0
Lesser Slave Lake	BOTO	Overall	40	0.05	0	0	40	5.2	3.3-7.5	17.5	5.0-50.0
Aug 2 – Aug 16	WOFR	Adult	382	0.48	0	0	382	2.9	1.5-5.2	2.9	2.0-20.0
(800 trap nights)	WOFR	YOY	203	0.25	0	0	203	2.4	1.0-4.3	1.3	0.5-2.4
	WOFR	Overall	621	0.78	0	0	621	2.7	1.0-5.2	2.3	0.5-20.0
Saskatoon Island	BCFR	Overall	2	0.005	0	0	2	1.6	1.5-1.6	1.5	1.4-1.5
July 28 - Aug 16	WOFR	Overall	156	0.39	0	0	156	3.2	1.8-5.7	5.9	1.4-20.0
(400 trap nights)											
Cypress Hills	NLFR	Overall	71	0.044	0	0	71	4.3	3.4-9.2	9.9	3.0-60.0
July 3 – Aug 30	TISA	Overall	4	0.0025	0	0	4	9.5	8.1-10.6	29.4	20.5-37.5
(1629 trap nights)											
	BCFR	Overall	16	0.01	1	0	15	1.7	1.4-2.7	0.4	0.3-1.0
Moonool	BOTO	Overall	23	0.015	2	10	=	4.3	1.4-7.0	12.2	0.3-45.5
Inly A And 12	WOFR	Adult	86	0.062	26	19	53	4.1	0.7-5.1	8.8	2.5-18.0
July 4 – Aug 12 (1571 tran nights)	WOFR	Juvenile	46	0.029	0	0	46	2.9	1.5-3.9	2.8	1.5-8.0
(1571 trap mgms)	WOFR	YOY	838	0.53	0	0	838	2	1.0-2.8	6.0	2.2
	WOFR	Overall	982	0.63	26	19	937	2.3	0.7-5.1	1.7	0.3-18.0

				# Per		2		Snout	Snout-to-Vent	-	
RANA SITE	Species*	Age**	Total#	Trap		Sex		Leng	Length (cm)	<u>-</u>	Mass (g)
				Night	M	Ŧ	Unk.	Mean	Range	Mean	Range
	NLFR	Overall	71	0.014	0	0	71	4.3	3.4-9.2	9.9	3.0-60.0
	TISA	Overall	4	8E-04	0	0	4	9.5	8.1-10.6	29.4	20.5-37.5
	BCFR	Overall	32	90000	1	0	31	1.8	1.4-3.6	6.0	0.4-1.5
	BOTO	Overall	91	0.018	4	10	77	5.3	2.2-7.3	18.4	12.2-26.0
	CSFR	Overall	1	2E-04	0	0	1	8.1	8.1	69.4	69.4
Totals	LTSA	Adult	9	0.001	4	2	0	6.4	5.6-7.5	5.6	3.8-8.4
(5154 trap nights)	LTSA	YOY	10	0.002	0	0	10	4.1	3.7-4.5	1.7	1.2-2.0
	LTSA	Overall	173	0.034	4	2	10	4.1	3.7-7.5	2.2	0.98-3.5
	WOFR	Adult	501	0.097	33	25	443	3.9	0.7-5.5	8.2	1.7-20.0
	WOFR	YOY	1048	0.2	0	0	1048	2.2	1.0-4.3	1.3	0.3-2.2
	WOFR	Juvenile	46	0.009	0	0	46	2.9	1.5-3.9	2.8	1.5-8.0
	WOFR	Overall	1989	0.39	33	25	1537	2.8	0.7-5.7	4.2	0.3-20.0

*BCFR (Boreal Chorus Frog), BOTO (Boreal Toad), CSFR (Columbia Spotted Frog), LTSA (Long-Toed Salamander), WOFR (Wood Frog), NLFR (Northern Leopard Frog), TISA (Tiger Salamander)

**Adult LTSA and WOFR were 2.0g or more, whereas YOY (young-of-the-year) LTSA and WOFR were <2.0g

Table 3. Amphibian species observations from all pond surveys in 2003.

	Total #		Num	ber of por	ids in whi	Number of ponds in which species* were observed	* were obs	served	
RANA Site	of Ponds Surveyed	BCFR	BOTO	CATO	CSFR	LTSA	NLFR	TISA	WOFR
Cypress Hills	15	3	0	0	0	0	7	2	0
Hinton	100	7	25	0	1	58	0	0	78
Jasper National Park	29	0	2	0	1	13	0	0	0
Kananaskis	82	0	19	0	14	17	0	0	28
Lesser Slave Lake	6	8	1	0	0	0	0	0	4
Meanook	0	0	0	0	0	0	0	0	0
Saskatoon Island	8	8	3	0	0	1	0	0	8
Totals	243	26	51	0	16	06	7	7	120

^{*}BCFR (Boreal Chorus Frog), BOTO (Boreal Toad), CATO (Canadian Toad), CSFR (Columbia Spotted Frog), LTSA (Longtoed Salamander), NLFR (Northern Leopard Frog), TISA (Tiger Salamander), WOFR (Wood Frog).

Table 4. Summary of educational/interpretive presentations given to the public from all Alberta RANA sites in 2003.

5.0 DISCUSSION

The types of species captured and observed at RANA sites in 2003 were consistent with data from previous years. Wood frogs continue to be the most ubiquitous species, being widely distributed across the province and the most abundant species captured. With the exception of Hinton, wood frog capture rates were higher than in all previous years. However, evidence of wood frog breeding was common in the Hinton area. The reason for the apparent population increase is unknown, but is not unexpected due to natural population fluctuations.

Long-toed salamander population persistence continues to be fairly stable in Hinton. Population persistence in Kananaskis is similar to last year, but relative to surveys in the mid-1990's, population persistence has gone down (refer to Wilkinson and Hanus 2003). Increased human activity and development may be contributing to this apparent reduction in breeding ponds. However, trapping data from the Kananaskis RANA pond, a relatively undisturbed site, suggest a healthy local breeding population. The Hinton site had unusually low numbers of young-of-the-year relative to the number of eggs laid, which may be a result of extremely low water levels at the trapping pond, contributing to increased competition and predation. Ongoing monitoring is needed to understand trends. Expanded surveys and reported observations from northwestern Alberta revealed the presence of long-toed salamanders near the town of Peace River, representing a range extension. In addition to observations near Fairview, there was an unconfirmed observation south of the Peace River. It is important to continue expanding surveys in northern Alberta to determine the range of the long-toed salamander population.

There were two captures of Columbia spotted frogs in 2003. Columbia spotted frogs are rarely encountered in the RANA program because they are found in the Rocky Mountains at higher elevations, where survey sites are under represented. Backcountry surveys and coordinated efforts with Parks are needed to effectively inventory Columbia spotted frogs. Boreal chorus frogs tend to be absent in the Rocky Mountains and uncommon in the foothills; however, they are usually at high numbers at Lesser Slave Lake. This was a below average year for boreal chorus frogs at Lesser Slave Lake, which may be due to the population explosion of wood frogs. Although boreal chorus frogs are a fairly common species, we lack information on their population trends and should not assume their populations are stable.

The boreal toad was the third-most abundant species captured in 2003, found in four of the six RANA sites. Capture rates were generally similar to previous years, recognizing that there have been considerable fluctuations between years. There is evidence of boreal toad population declines in other jurisdictions, so it is important to continue monitoring their population trends in Alberta. Similarly, it is important to expand survey efforts within Canadian toad range because there has been only one record throughout the history of the RANA program. The toad was captured in 1998 at Lesser Slave Lake, which is at the western edge of Canadian toad distribution in Alberta (the range is roughly limited to the eastern half of the province). Neither Meanook nor Beaverhill Lake RANA sites have recorded Canadian toads, despite being located in Canadian toad range.

Northern leopard frogs were only observed in the Cypress Hills area, which is expected due to their known range. Given their current status as 'threatened' it is encouraging that there was evidence of breeding at the trapping pond. Pitfalls traps were successfully modified to be deeper and wider, improving capture efficiency. Seven of the 15 ponds surveyed at Cypress Hills had northern leopard frogs, of which five showed breeding activity. The number of tiger salamanders captured and observed in Cypress Hills was low, and as such was similar to previous years. Monitoring of a previously identified highway crossing of tiger salamanders west of Calgary did not yield any observations (other than wood frogs; Rose 2004). Surveys of nearby ponds and discussions with locals did not yield tiger salamander observations. Our understanding of tiger salamander mass movements is minimal, and it raises the question of whether heavily used roads pose a threat to migrating amphibians (see Rose 2004 for details).

Although the RANA program has operated for seven years, this is still a relatively short time period from which to draw conclusions about amphibian population trends due to their stochasticity. With improved consistency and continued monitoring we are beginning to generate an important long-term data set about amphibian distribution and population trends in Alberta. Currently, RANA data provide information on amphibian species presence and distribution, which also serve as a warning that a species may potentially be in decline due to its absence, or that certain breeding sites may be vulnerable. The data provide valuable information on timing of breeding as well as checking for possible deformities or disease. In addition, considerable life history information has been gathered. The variability in the numbers of species caught annually reinforces the need to conduct long-term monitoring.

Few amphibian species in Alberta are considered secure and wetland habitat is under threat of alteration or destruction. Maintenance of RANA sites, especially those sites in key habitats with long monitoring histories, is strongly recommended. Aside from amphibian monitoring, the RANA program provides extensive public education about amphibians and wetland conservation, which is arguably as important as monitoring. In recognition of global amphibian declines and national efforts to track amphibian population trends, Alberta's maintenance of the RANA program is critical.

6.0 MANAGEMENT IMPLICATIONS AND FUTURE RESEARCH

Management recommendations are limited given the relatively short period of time RANA has been operating. The primary recommendation is preservation of wetlands (*i.e.* prevent draining and contamination), particularly those known to have breeding populations of species at risk, and they should be carefully managed to ensure persistence of amphibian populations. Fish stocking can also have a detrimental effect on amphibians, particularly long-toed salamanders (refer to Rose 2004). We must balance the needs of amphibians with recreational activities. Minimizing potentially destructive activities around ponds is an additional consideration. On crown land, protective notations have been applied to create buffers around some breeding ponds. It is important to maintain communication with landowners and people who participate in activities around critical amphibian ponds in order to foster a sense of stewardship and appreciation for wetland ecosystems, and discourage collection of amphibians. Signage and education programs should be integrated with management activities. Future survey efforts should

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8.0 APPENDICES

Appendix 1. Summary of historic amphibian captures at each RANA site.

A -Beaverhill				
Year	# Trap Nights	BCFR*	TISA*	WOFR*
		Total # caught	Total # caught	Total # caught
		(# caught per trap night)	(# caught per trap	(# caught per trap
			nignt)	mgm)
1998	2073	17 (0.008)	1 (0.000)	182 (0.089)
1999	1257	1 (0.001)	1 (0.001)	111 (0.088)
2000	186	2 (0.011)	0 (0.000)	14 (0.075)
3CED (Borest Chorn	IS From TISA (Tine	BCER (Boreal Chorus Eron) TISA (Tiger Salamander) WOFR (Wood Frog)	Vond Froe	

*BCFR (Boreal Chorus Frog), TISA (Tiger Salamander), WUFK (WOOU Frog).

B - Cypress Hills Provincial Park				
Year	# Trap Nights	BCFR*	NLFR*	TISA*
		Total # caught	Total # caught	Total # caught
		(# caught per trap night)	(# caught per trap	(# caught per trap night)
1998	180	2 (0.011)	5 (0.028)	31 (0.172)
1999	927	4 (0.004)	20 (0.022)	12 (0.013)
2000	1440	2 (0.001)	19 (0.013)	15 (0.010)
2001	N/A	N/A	N/A	N/A
2002	N/A	N/A	N/A	N/A
2003	2629	2 (0.00076)	71 (0.027)	10 (0.0038)

*BCFR (Boreal Chorus Frog), NLFR (Northern Leopard Frog), TISA (Tiger Salamander).

# Trap Nights	BOTO*		
# Trap Nights	BOTO*		
742	Total # caught	LTSA*	WOFR*
247	0	Total # caught	Total # caught
7/13	(# caught per trap night)	(# caught per trap night)	(# caught per trap night)
	44 (0.058)	135 (0.180)	346 (0.468)
2001 1072 1	13 (0.012)	161 (0.150)	69 (0.064)
2002 1646 2	29 (0.018)	218 (0.132)	175 (0.106)
2003 946 1	10 (0.011)	108 (0.11)	54 (0.057)

*BOTO (Boreal Toad), LTSA (Long-toed Salamander), WOFR (Wood Frog).

D – Nananaskis and area					
Year	# Trap Nights	BOTO*	CSFR*	LTSA*	WOFR*
		Total # caught	Total # caught	Total # caught	Total # caught
		(# caught per trap night)			
1998	1068	33 (0.031)	7 (0.007)	186 (0.174)	61 (0.057)
1999	383	11 (0.029)	2 (0.005)	10 (0.026)	12 (0.031)
2000	522	3 (0.006)	0 (0.000)	6 (0.011)	3 (0.006)
2001	484	11 (0.023)	1 (0.002)	34 (0.070)	8 (0.017)
2002**	1363	39 (0.029)	3 (0.002)	465 (0.341)	213 (0.156)
2003	1072	27 (0.03)	1 (0.0009)	283 (0.26)	218 (0.2)

* BOTO (Boreal Toad), CSFR (Columbia Spotted Frog), LTSA (Long-Toed Salamander), WOFR (Wood Frog).

**In 2002 a new RANA trapping site was established, Kuhn's Pond, due to dry-up of old pond for 3 consecutive years.

E-Lesser Slave L	E -Lesser Slave Lake Provincial Park	BCFR*	BOTO*	WOFR*
Vear	# Trap Nights	Total # caught	Total # caught	Total # caught
	0	(# caught per trap night)	(# caught per trap night)	(# caught per trap night)
1997	724**	8 (0.011)	7 (0.010)	73 (0.101)
***8661	3456	5 (0.001)	23 (0.007)	33 (0.001)
1999	3312	57 (0.017)	114 (0.034)	119 (0.036)
2000	3216	26 (0.008)	84 (0.026)	52 (0.016)
2001	840	29 (0.035)	817 (0.973)	83 (0.099)
2002	1680	299 (0.178)	156 (0.093)	196 (0.117)
2003	1360	52 (0.0038)	67 (0.049)	809 (0.59)

*BCFR (Boreal Chorus Frog), BOTO (Boreal Toad), WOFR (Wood Frog).

**The number of trap nights is an estimate due to difficulties interpreting the number of trap malfunctions in original data.

***One Canadian toad was captured in 1998.

F-Meanook Biological Research Station	gical Research	BCFR*	BOTO*	WOFR*
Year	# Trap Nights	Total # caught	Total # caught	Total # caught
	0	(# caught per trap night)	(# caught per trap night)	(# caught per trap night)
1997	518	4 (0.008)	3 (0.006)	193 (0.387)
1998	755	4 (0.005)	343 (0.454)	277 (0.367)
1999	630	2 (0.003)	7 (0.011)	23 (0.037)
2000	2090	6 (0.003)	125 (0.06)	36 (0.017)
2001	644	4 (0.006)	8 (0.012)	316 (0.49)
2002	714	1 (0.001)	12 (0.017)	65 (0.091)
2003	1571	16 (0.01)	23 (0.015)	982 (0.63)

*BCFR (Boreal Chorus Frog), BOTO (Boreal Toad), WOFR (Wood Frog).

J-Saskatoon Islan	G-Saskatoon Island Provincial Park	BCFR*	BOTO*	WOFR*
Vear	# Tran Nights	Total # caught	Total # caught	Total # caught
		(# caught per trap night)	(# caught per trap night)	(# caught per trap night)
6661	1070	9 (0.008)	0 (0.000)	128 (0.120)
2000	1081	17 (0.016)	2 (0.002)	44 (0.041)
2001	966	5 (0.005)	3 (0.003)	74 (0.074)
2002	086	4 (0.004)	0 (0.000)	165 (0.168)
2003	808	11 (0.014)	0 (0.000)	177 (0.22)

*BCFR (Boreal Chorus Frog), BOTO (Boreal Toad), WOFR (Wood Frog).

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